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(54) Title: CYCLIC AGONISTS AND ANTAGONISTS OF C52 RECEPTORS AND G PROTEIN-COUPLED RECEPTORS

#### (57) Abstract

This invention relates to novel cyclic compounds which have the ability to modulate the activity of G protein-coupled receptors. The invention provides both agonists and antagonists. In preferred embodiments, the invention provides cyclic peptidic and cyclic or non-cyclic non-peptidic antagonists or agonists of C5a. The compounds of the invention are both potent and selective, and are useful in the treatment of conditions mediated by G protein-coupled receptors, especially conditions mediated by overexpression or underregulation of C5a, such as a variety of inflammatory conditions.

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## CYCLIC AGONISTS AND ANTAGONISTS OF C5a RECEPTORS AND G PROTEIN-COUPLED RECEPTORS

This invention relates to novel cyclic compounds

which have the ability to modulate the activity of G

protein-coupled receptors. The invention provides both

agonists and antagonists. In preferred embodiments, the

invention provides cyclic peptidic and cyclic or non-cyclic

non-peptidic antagonists or agonists of C5a. The compounds

of the invention are both potent and selective, and are

useful in the treatment of a variety of inflammatory

conditions.

#### BACKGROUND OF THE INVENTION

Activation of human complement, a system of plasma proteins involved in immunological defence against infection and injury, contributes significantly to the pathogenesis of numerous acute and chronic diseases. In particular, the complement protein C5a has been extensively investigated. For general reviews, see Whaley (1987), and Sim (1993). Table 1 provides a summary of known roles of C5a in disease.

During host defence, the complement system of plasma proteins initiates inflammatory and cellular immune responses to stimuli such as infectious organisms 25 (bacteria, viruses, parasites), chemical or physical injury, radiation or neoplasia. Complement is activated through a complex cascade of interrelated proteolytic events that produce multiple bioactive peptides, some of 30 which (eg. anaphylatoxins C3a and C5a) interact with cellular components to propagate inflammatory processes. Complement activation, either by the classical pathway, after antigen-antibody (Ag/Ab) binding, or by the antibodyindependent alternate pathway, ends with a terminal 35 sequence in which protein C5 is proteolytically cleaved by C5 convertase to C5a and C5b. The latter facilitates assembly of a "membrane attack complex" that punches holes

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in membranes of target cells such as bacteria, leading to leakage, lysis and cell death. Steps in the cascade are tightly regulated to avoid stepwise amplification of proteolysis by sequentially formed proteases. If these regulatory mechanisms become inefficient, protracted activation of complement can result, causing enhanced inflammatory responses as in autoimmune diseases.

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Although the broad features of the complement system and its activation are known, mechanistic details remain poorly understood. A principal and very potent mediator of inflammatory responses is the plasma glycoprotein C5a, which interacts with specific surface receptors (C5aR) on mast cells, neutrophils, monocytes, macrophages, non-myeloid cells, and vascular endothelial cells (Gerard and Gerard, 1994). C5aR is a G protein-coupled receptor with seven transmembrane helices (Gerard and Gerard, 1991). This receptor is one of the rhodopsin superfamily of GTP-linked binding proteins, but differs from rhodopsin receptors in that the receptor and G protein are linked prior to activation.

G protein-coupled receptors are prevalent throughout the human body, comprising approximately 80% of known cellular receptor types, and mediate signal transduction across the cell membrane for a very wide range of endogenous ligands. They participate in a diverse array of physiological and pathophysiological processes, including, but not limited to those associated with cardiovascular, central and peripheral nervous system, reproductive, metabolic, digestive, immunoinflammatory, and growth disorders, as well as other cell-regulatory and proliferative disorders. Agents, both agonists and antagonists, which selectively modulate functions of G protein-coupled receptors have important therapeutic applications.

C5a is one of the most potent chemotactic agents known, and recruits neutrophils and macrophages to sites of injury, alters their morphology; induces degranulation;

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summarised in Table 1.

increases calcium mobilisation, vascular permeability (oedema) and neutrophil adhesiveness; contracts smooth muscle; stimulates release of inflammatory mediators (including histamine, TNF-α, IL-1, IL-6, IL-8, prostaglandins, leukotrienes) and lysosomal enzymes; promotes formation of oxygen radicals; and enhances antibody production (Gerard and Gerard, 1994).

Overexpresion or underregulation of C5a is implicated in the pathogenesis of immunoinflammatory conditions such as rheumatoid arthritis, adult respiratory distress syndrome (ARDS), systemic lupus erythematosus, tissue graft rejection, ischaemic heart disease, reperfusion injury, septic shock, psoriasis, gingivitis, atherosclerosis, Alzheimer's disease, lung injury and extracorporeal post-dialysis syndrome, and in a variety of other conditions, as

Table 1 The Role of C5a in Disease

| Condition/disease      | CSa                                     | C5aR                                    |   |
|------------------------|---|---|---|
|                        | levels                                  | expression                              | Details                                       |
| allergy                | <b>+</b> +                              |   | allergen challenge leads to nasal symptoms    |
|                        |   |   | and increased C5a levels                      |
| Alzheimer's disease ++ |   | ++                                      | up-regulation of the receptor in reactive     |
|                        |   |   | astrocytes, microglia and endothelial cells   |
|                        |   |   | in the CNS, complement system activated by 8- |
|                        | *************************************** |   | amyloid                                       |
| ARDS/respiratory       | <b>+</b><br>+                           |   |   |
| distress               |   |   |   |
| V.                     | ++                                      |   | levels highest just prior to ocular attack    |
| bronchial asthma       | ++                                      | *************************************** |   |
| capillary leak         | +<br>+                                  |   |   |
| syndrome               | *************************************** |   |   |

| Condition/disease   | C5a           | C5aR                                    | 1       |
|---------------------|---------------|---|---|
| 9.1                 | levels        | expression                              | Decails                                       |
| chronic lung        | <b>‡</b>      |   | Increased C5a levels in pulmonary effluent    |
| disease             |               |   | fluid from mechanically ventilated infants    |
| -                   |               |   | with chronic lung disease                     |
| Churg-Strauss       |               |   | hypersensitivity of granulocytes to C5a       |
| cystic fibrosis     |               | ****                                    | generation of C5a/effects on PMNs             |
| decompression       | <b>+</b><br>+ |   | increased C5a levels during saturation diving |
| stress              |               |   |   |
| diabetes type I     | ++            |   | C5a generated during onset; circulating       |
|                     |               |   | monocytes in newly diagnosed type 1 diabetes  |
|                     |               |   | patients are activated                        |
| Familiai            |               |   | lack of C5a inactivator                       |
| Mediterranean fever |               | *************************************** |   |
| Guillain-Barre      | +             | *************************************** | CSF levels elevated                           |
| ischaemic disease   |               |   | migration of monocytes into myocardium after  |
| states/ myocardial  |               |   | reperfusion. Damage prevented with sCR1       |
| infarct             |               |   |   |

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| Condition/disease      | CSa                                     | CS&R                                    |  |
|------------------------|---|---|--|
|                        | levels                                  | expression                              | Details                                    |
| Kimura's disease       |   |   | r up-regulates t                           |
|                        |   |   | PMNs to C5a                                |
| Multiple Sclerosis     | •                                       | ++                                      | xpressio                                   |
| •                      |   |   | macrophages in acute and chronic MS and    |
|                        |   |   | fibrous astrocytes in chronic MS           |
| Meningitis             |   |   | C5a induces experimental meningitis; PMN   |
|                        | 1 | *************************************** | accumulation seen in the CSF               |
| pancreatitis ++        | ++                                      |   |  |
| ဟ                      | +<br>+                                  | 1                                       | C5a generated via complement activation by |
| syndrome               |   |   | tubing material, C5aR levels decreased on  |
|                        |   |   | PMNs & monocytes in chronic state          |
| preeclampsia/ HELLP ++ | +<br>+                                  | *************************************** | C5a levels elevated at delivery            |
| psoriasis              |   |   | C5a levels high in scales                  |
| ŭ                      | ++                                      |   | inhibited by C5 antibody                   |
| retinitis              | ++                                      |   | C5a detected in vitreous humor             |

| Condition/disease | CSa                                     | CSaR                                    | •   |
|-------------------|---|---|---|
| le                | levels                                  | expression                              | xpression   |
| Rheumatoid        | +++++++++++++++++++++++++++++++++++++++ |   | elevated concentration of C5a found in                |
| arthritis         | 10                                      | *************************************** | arthritis synovial fluid (5-fold) and plasma (3-fold) |
| Severe congenital |   |   |   |
| neutropenia       | *************************************** | *************************************** | neutropenia   |
| transplant/graft  | †<br>+                                  |   | monoclonal antibodies block the damage                |
| rejection         |   |   | seen with xenogenic transplant; increased             |
|                   |   |   | levels of C5a seen in the plasma and                  |
|                   |   |   | urine of patients with renal graft                    |
|                   |   |   | rejection   |

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New agents which limit the pro-inflammatory actions of C5a have potential for inhibiting chronic inflammation, and its accompanying pain and tissue damage. For these reasons, molecules which prevent C5a binding to its receptors are useful for treating chronic inflammatory disorders driven by complement activation. Importantly, such compounds provide valuable new insights to mechanisms of complement-mediated immunity.

In another context, agonists of C5a receptors or other G protein-coupled receptors may also be found to have therapeutic properties in conditions either where the G protein-coupled receptor can be used as a recognition site for drug delivery, or where triggering of such receptors can be used to stimulate some aspect of the human immune system, for example in the treatment of cancers, viral or parasitic infections.

One approach to the development of agonists or antagonists of C5a is through receptor-based design, using knowledge of the three-dimensional structures of C5a, its receptor C5aR, and the interactions between them. The structure of the receptor is unknown. The solution structure of human C5a, a 74 amino acid peptide that is highly cationic and N-glycosylated with a 3 kDa carbohydrate at Asn64, has been determined and is essentially a 4-helix bundle. The C-terminal end (residues 65-74, C5a65-74) was found to be unstructured (Zuiderweg et al, 1989) and this conformational flexibility in the C-terminus has made structure-function studies extremely difficult to interpret.

C5a has a highly ordered N-terminal core domain (residues 1-64; C5a<sub>1</sub>-64), consisting of a compact antiparallel 4-helix bundle (residues 4-12, 18-26, 32-39, 46-63) connected by loops (13-17, 27-31, 40-45), and further stabilised by 3 disulphide bonds (C21-Cys47, Cys22-35 Cys54, Cys34-Cys55).

Although the structure of the C5a receptor, C5aR, is unknown, the C5a-binding subunit of human monocyte-

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derived C5aR has been cloned and identified as a G proteincoupled receptor with transmembrane helices (Gerard and Interactions between C5a and C5aR have been Gerard, 1991). the subject of many investigations which, in summary, suggest that C5a binds via a two-site mechanism in which the N-terminal core domain of C5a is involved in receptorrecognition and binding, while the C-terminus is This mechanism is responsible for receptor activation. illustrated schematically in Figure 1. The C-terminal "effector" region alone possesses all the information 10 necessary for signal transduction, and is thought to bind in the receptor's interhelical region (Siciliano et al, 1994; deMartino et al, 1995).

An N-terminal interhelical positively-charged
region of C5a is responsible for receptor recognition and
binding, and binds to a negatively-charged extracellular
domain of C5aR (site 1), while the C-terminal "effector"
region of C5a is thought to bind with the interhelical
region of the receptor (site 2), and is responsible for
receptor activation leading to signal transduction
(Siciliano et al, 1994).

Numerous short peptide derivatives of the Cterminus of C5a have been found to be agonists of C5a
(Kawai et al, 1991; Kawai et al, 1992; Kohl et al, 1993;
Drapeau et al, 1993; Ember et al, 1992; Sanderson et al,
1994; Sanderson et al, 1995; Finch et al, 1997; Tempero et
al, 1997; Konteatis et al, 1994; DeMartino et al, 1995).
The structures of some of these agonists are shown in Table
2 below (compounds 1-6). High molecular weight polypeptide
inhibitors of the action of C5a at its receptor, such as
monoclonal antibodies to the C5a receptor, are also known
(Morgan et al, 1992).

A small molecule, N-methylphenylalanine-lysine-proline-D-cyclohexylalanine-tryptophan-D-arginine (7, MeF- K-P-dCha-W-r), is a full antagonist of the C5a receptor with no agonist activity when tested on isolated cellular membranes (Konteatis et al, 1994) or intact whole cells.

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This hexapeptide was developed by modifications of the agonist NMe-F-K-P-dCha-L-r, in which the molecule was progressively substituted at leucine residues with substituents of increasing size (Cha, F, Nph and W). This had the effect of reducing agonist activity. Receptor-binding assays, performed on isolated human neutrophil membranes, showed that the antagonist had only 0.04% relative affinity of C5a for the receptor (Konteatis et al, 1994). A key feature of these reports is the definition of the binding of 7 to the C5a receptor. These authors state that the C-terminal arginine is essential for receptor binding and antagonist activity. This is also the case in all the reports of agonist activity by small peptide analogues of the C-terminus of C5a. However, for the antagonist 7, the authors go further and state that "the C-terminal carboxylate is an essential"

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"the C-terminal carboxylate is an essential requirement for antagonist activity and receptor binding."

They proposed that the requirement of the carboxylate is probably the result of its specific 20 interaction with an arginine (Arg 206) in the receptor (De Martino et al, 1995). This idea was supported by a great reduction in receptor-affinity for an analogue of 7 in which the D-arginine  $(NH_2-CH(CO_2H)-(CH_2)_3NHC(:NH)NH_2)$  was replaced by agmatine (NH2-CH2-(CH2)3NHC(:NH)NH2). 25 summary, De Martino et al claim that the D-arginine interacts via its guanidinium side chain with a negativelycharged amino acid side chain in the receptor. A second interaction between the negatively-charged C-terminal carboxylate of 7 and a positively-charged side chain 30 residue in the receptor is also thought to occur.

We have now determined the solution structure of this hexapeptide 7 and several analogues, and have surprisingly found that in fact a terminal carboxylate group is not required for binding to C5aR or for antagonist activity, and that instead an unusual hitherto unrecognised structural feature, a turn conformation, is responsible for

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C5a antagonist or agonist binding and activity. hexapeptide and several new structurally related antagonists have been examined for both their receptorbinding affinities and antagonist activity, using intact polymorphonuclear (PMN) cells. Our results show the hitherto unknown specific structural requirement for the binding of C5a antagonists or agonists to the C5a receptor, which we believe to be common to ligands for the G proteincoupled receptor family. Our establishment of this specific structural requirement has enabled us to design 10 and develop improved molecular probes of the complement system and of C5a-based drugs, and to design small molecules that target other G protein-coupled receptors, which are becoming increasingly recognised as important drug targets due to their crucial roles in signal 15 transduction (G protein-coupled Receptors, IBC Biomedical Library Series, 1996).

Thus our results have enabled us to design constrained structural templates which enable hydrophobic groups to be assembled into a hydrophobic array for interaction with a G protein-coupled receptor, for example at Site 2 of the C5a receptor illustrated in Figure 1. Such templates or scaffolds, which may be cyclic or acyclic, have not heretofore been suggested for modulators of the activity of C5a receptors or other G protein-coupled receptors.

#### SUMMARY OF THE INVENTION

The invention provides cyclic and non-cyclic modulators of the activity of G-coupled-protein receptors.

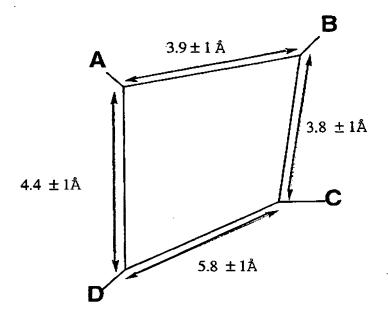
According to a first aspect, the invention provides a compound which is an antagonist, of a G-coupled-protein receptor, which has no agonist activity, and which has a structure adapted to provide a framework of

35 approximate dimensions as follows:

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#### Structure I



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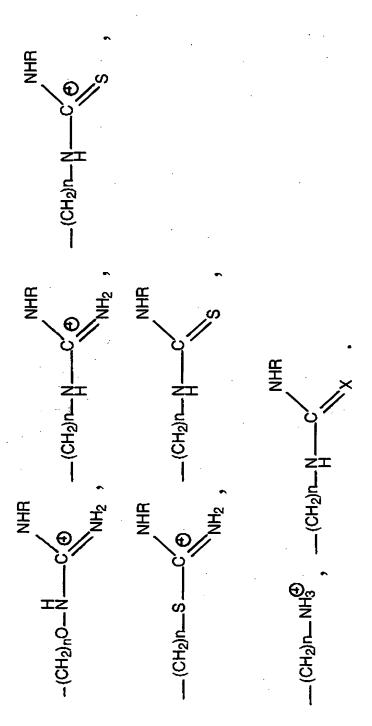
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where the numerals refer to distances between  $C_\alpha$  carbons of amino acids or their analogues or derivatives, and A, B, C and D are not necessarily on adjacent amino acids, or analogues or derivatives thereof; and

where the critical amino acid side chains are designated by A, B, C and D, or are as defined below;

A is any common or uncommon, basic, charged amino acid side chain which serves to position a positively charged group in this position, including, but not limited to the following side chains and other mimetics of arginine side chains:

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where

X is H, NCN, NNO2, CHNO2 or NSO2NH2;

n is an integer from 1 to 4, and

R is H or an alkyl, aryl, CN, NH2 or OH group .

B is any common or uncommon aromatic amino acid side chain which serves to position an aromatic side-chain group in this position, including but not limited to the indole, indole methyl, benzyl, phenyl, naphthyl, naphthyl methyl, cinnamyl group, or any other derivatives of these aromatic groups;

C is any common or uncommon hydrophobic amino acid side chain which serves to position any alkyl, aromatic or other group in this position, including, but not limited to D- or L-cyclohexyl alanine (Cha), leucine, valine, isoleucine, phenylalanine, tryptophan, or methionin

D is any common or uncommon aromatic amino acid which serve to position an aromatic side-chain in this position, and has the structure:

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where Z includes but is not limited to indole, indole methyl, benzyl, benzene, naphthyl, naphthyl methyl, or any other derivatives of these aromatic groups, and

R is H or any alkyl, aromatic, acyl or aromaticacyl group including, but not limited to methyl, ethyl, propyl, butyl, -CO-CH<sub>2</sub>CH<sub>3</sub>, -CO-CH<sub>2</sub>CH<sub>3</sub>, -CO-CH<sub>2</sub>CH<sub>3</sub>, -CO-CH<sub>2</sub>Ph, or -CO-Ph.

Preferably the G protein-coupled receptor is a C5a receptor.

Other cyclic or non-cyclic molecules, which may be peptidic or non-peptide in nature, can similarly be

envisaged to support groups such as A, B, C and D for interaction with a C5a receptor or other G protein-coupled receptor.

In one preferred embodiment, the compound has antagonist activity against C5aR, has no C5a agonist activity, and has the general formula:

#### Structure II

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where A is H, alkyl, aryl, NH<sub>2</sub>, NHalkyl, N(alkyl)<sub>2</sub>,
15 NHaryl or NHacyl; OH, Oalkyl, Oaryl.

B is an alkyl, aryl, phenyl, benzyl, naphthyl or indole group, or the side chain of a D- or L-amino acid selected from phenylalanine, homophenylalanine, tryptophan, homotryptophan, tyrosine, and homotyrosine;

C is the side chain of a D-, L- or homo-amino acid selected from the group consisting of proline, alanine, leucine, valine, isoleucine, arginine, histidine, aspartate, glutamate, glutamine, asparagine, lysine, tyrosine, phenylalanine, cyclohexylalanine, norleucine, tryptophan, cysteine and methionine;

D is the side chain of a D- or L-amino acid selected from the group consisting of cyclohexylalanine, homocyclohexylalanine, leucine, norleucine, homoleucine, homonorleucine and tryptophan;

E is the side chain of a D- or L-amino acid selected from the group consisting of tryptophan and homotryptophan;

F is the side chain of a D- or L-amino acid selected from the group consisting of arginine, homoarginine, lysine and homolysine; and

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X is  $-(CH_2)_nNH-$  or  $(CH_2)_n-S-$ , where n is an integer of from 1 to 4, preferably 2 or 3,  $-(CH_2)_2O-$ ,  $-(CH_2)_3O-$ ,  $-(CH_2)_3-$ ,  $-(CH_2)_4-$ , or  $-CH_2COCHRNH-$ , where R is the side chain of any common or uncommon amino acid.

For the purposes of this specification, the term "alkyl" is to be taken to mean a straight, branched, or cyclic, substituted or unsubstituted alkyl chain of 1 to 6, preferably 1 to 4 carbons. Most preferably the alkyl group is a methyl group. The term "acyl" is to be taken to mean a substituted or unsubstituted acyl of 1 to 6, preferably 1 to 4 carbon atoms. Most preferably the acyl group is acetyl. The term "aryl" is to be understood to mean a substituted or unsubstituted homocyclic or heterocyclic aryl group, in which the ring preferably has 5 or 6 members.

A "common" amino acid is a L-amino acid selected from the group consisting of glycine, leucine, isoleucine, valine, alanine, phenylalanine, tyrosine, tryptophan, aspartate, asparagine, glutamate, glutamine, cysteine, methionine, arginine, lysine, proline, serine, threonine and histidine.

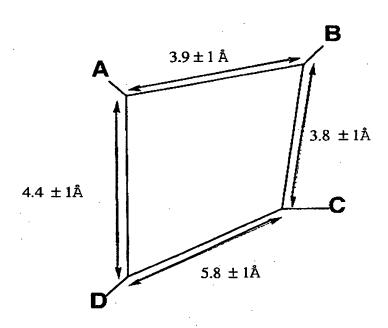
An "uncommon" amino acid includes, but is not restricted to, D-amino acids, homo-amino acids, N-alkyl amino acids, dehydroamino acids, aromatic amino acids (other than phenylalanine, tyrosine and tryptophan), ortho-, meta- or para-aminobenzoic acid, ornithine, citrulline, norleucine,  $\gamma$ -glutamic acid, aminobutyric acid and  $\alpha, \alpha$ -disubstituted amino acids.

For the purposes of this specification it will be clearly understood that the word "comprising" means

"including but not limited to", and that the word "comprises" has a corresponding meaning.

According to a second aspect, of the invention provides a compound which is an agonist of G protein-coupled receptors, and which has structure III

#### Structure III



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where the numerals refer to distances between  $C_{\alpha}$  carbons of amino acids or their analogues or derivatives, and A, B, C and D are not necessarily on adjacent amino acids, or analogues or derivatives thereof; and

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where B is a non-aromatic amino acid, and is preferably the D- or L-form of alanine, leucine, valine, norleucine, glutamic acid, aspartic acid, methionine, cysteine, isoleucine, serine or threonine,

and A, C and D are as defined above.

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Preferably the compound is of structure IV,

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#### Structure IV

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where E is any amino acid other than tryptophan and homotryptophan, for example D- or L- forms of alanine, leucine, valine, norleucine, phenylalanine, glutamic acid, aspartic acid, methionine, cysteine, isoleucine, serine, threonine. Preferably the compound is an agonist of C5a.

According to a third aspect, the invention provides a composition, comprising a compound according to the invention together with a pharmaceutically-acceptable carrier or excipient.

The compositions of the invention may be formulated for oral or parenteral use, but oral formulations are preferred. It is expected that most if not all compounds of the invention will be stable in the presence of digestive enzymes. Such stability can readily be tested by routine methods known to those skilled in the art.

Suitable formulations for administration by any desired route may be prepared by standard methods, for example by reference to well-known textbooks such as Remington; The Science and Practice of Pharmacy, Vol. II, 1995 (19<sup>th</sup> edition), A.R. Gennaro (ed), Mack Publishing Company, Easton, Pennsylvania, or Australian Prescription Products Guide, Vol. 1, 1995 (24<sup>th</sup> edition) J. Thomas (ed),

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Australian Pharmaceutical Publishing Company Ltd, Victoria, Australia.

In a fourth aspect, the invention provides a method of treatment of a pathological condition mediated by a G protein-coupled receptor, comprising the step of administering an effective amount of a compound of the invention to a mammal in need of such treatment.

Preferably the condition mediated by a G protein-coupled receptor is a condition mediated by a C5a receptor, and more preferably involves overexpression or underregulation of C5a. Such conditions include but are not limited to rheumatoid arthritis, adult respiratory distress syndrome (ARDS), systemic lupus erythematosus, tissue graft rejection, ischaemic heart disease, reperfusion injury, septic shock, psoriasis, gingivitis, atherosclerosis, Alzheimer's disease, lung injury and extracorporeal post-dialysis syndrome.

while the invention is not in any way restricted to the treatment of any particular animal or species, it is particularly contemplated that the compounds of the invention will be useful in medical treatment of humans, and will also be useful in veterinary treatment, particularly of companion animals such as cats and dogs, livestock such as cattle, horses and sheep, and zoo animals, including large bovids, felids, ungulates and canids.

The compounds may be administered at any suitable dose and by any suitable route. Oral administration is preferred because of its greater convenience and acceptability. The effective dose will depend on the nature of the condition to be treated, and the age, weight, and underlying state of health of the individual treatment. This will be at the discretion of the attending physician or veterinarian. Suitable dosage levels may readily be determined by trial and error experimentation, using methods which are well known in the art.

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## BRIEF DESCRIPTION OF THE FIGURES

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Figure 1 shows a diagrammatic representation of the two-site model for binding of C5a to its G protein-coupled receptor, C5aR. The black rods represent  $\alpha$ -helical regions, and the open cylinders represent the transmembrane helices. Sites 1 and 2 are indicated on the figure.

Figure 2 shows stacked plots of  $^{1}\text{H-NMR}$  spectra, showing time-dependent decay of amide NH resonances for Trp (8.10 ppm) and D-Cha (7.90 ppm) residues of **7** in d<sub>6</sub>-DMSO containing D<sub>2</sub>O after 10 minutes (bottom plot) and then 25, 40, 55, 70, 130, 190, 250, 385 and 520 minutes.

Figure 3 shows backbone C, N, O atoms of twenty lowest energy minimized NMR structures of 7 in  $d_6\text{-DMSO}$  at 24°C) .

Figure 4 shows a schematic representation of H-bonding in the structure of 7 from proton NMR spectra in d<sub>6</sub>-DMSO.

Figure 5a shows (a) receptor binding, as indicated by inhibition of binding of  $^{125}I-C5a$  to human PMNs by 7 ( $\bullet$ ); 8 ( $\Delta$ ); 9 ( $\triangle$ ); 12 (O).

Figure 5b shows C5a antagonist potency as inhibition of myeloperoxidase (MPO) release from human PMNs by : 7 ( $\blacksquare$ , n= 9) and 12 ( $\blacktriangle$ , n=4). Figure 5c shows C5aR binding and antagonist potencies of 7, 15 and 17.

A-C show the effect of increasing concentrations (top to bottom) of C5a antagonists inhibiting myeloperoxidase release in human PMNs (n=3 in A-C).

A: 7 at 0, 0.1, 0.3, 1.0 µM (top to bottom)

B: 15 at 0, 0.1, 0.03, 0.1  $\mu$ M (top to bottom)

C: 17 at 0, 0.01, 0.03, 0.1  $\mu$ M (top to bottom)

D: Comparative affinities for PMN C5qR receptor. Inhibition of binding of <sup>125</sup>I-C5a to human PMNs by **7** (top), **15** (middle), **17** (bottom). All data are means ± SEM.

Figure 6 shows receptor binding of cyclic C5a antagonists, as shown by inhibition of binding of <sup>125</sup>I-C5a to human PMNs (n=5).

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Figure 7 shows superimposed structures of 7 (light, NMR structure) and 12 (dark, computer modelled structure). Phe and Trp side chains are omitted from 12 for clarity.

Figure 8 shows inhibition of C5a-induced neutropenia in Wistar rats by the cyclic antagonist F-[OPdChaWR] given i.v. at 1mg/kg. Results shown from n=3 in each group, \*P<0.05 compared to C5a-treated group only. Results are expressed as mean ± SEM.

10 Figure 9 shows inhibition of LPS-induced neutropenia and changes in haematocrit induced by the cyclic antagonist F-[OPdChaWR] (0.03-10mg/kg, i.v., 10 min prior to lipopolysaccharide [LPS]) in Wistar rats.

Abscissa: time after LPS (1 mg/kg i.v. injection).

Ordinate: per cent change in haematocrit (A) value or level of circulating polymorphonuclear (PMN) leukocytes (B) compared to time zero.

Figure 10 shows inhibition of carrageenan-induced (Wistar) rat paw oedema by cyclic antagonist (3D35) AcF-[OPdChaWr] (1 mg/kg single dose i.p. given 30 min prior to carrageenan). Results shown from 4 rats /group, mean ± SEM. Ordinate: percent change in paw volume. Abscissa: time (mins) after carrageenan injection

#### 25 DETAILED DESCRIPTION OF THE INVENTION

The invention will now be described by way of reference only to the following general methods and experimental examples, and to the figures. Abbreviations used herein are as follows:

30 BOP benzotriazole-1-yl-oxy-tris-(dimethylamino)phosphonium hexafluorophosphate

D-Cha D-cyclohexylamine

DIPEA diisopropylethylamine

DMF dimethylformamide

35 DMSO dimethylsulphoxide

HBTU O-benzotriazole N',N',N',N'-tetramethyluronium hexafluorophosphate;

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LPS lipopolysaccharide

PMN polymorphonuclear granulocyte

RMSD root mean square deviation

RP-HPLC reverse phase-high performance liquid

5 chromatography

TFA trifluoroacetic acid;

Throughout the specification conventional singleletter and three-letter codes are used to represent amino acids.

#### General Methods

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Protected amino acids and resins were obtained from Novabiochem. TFA, DIPEA and DMF (peptide synthesis grade) were purchased from Auspep. All other materials were 15 reagent grade unless otherwise stated. Preparative scale reverse-phase HPLC separations were performed on a Vydac C18 reverse-phase column (2.2  $\times$  25 cm), and analytical reverse-phase HPLC separations were performed on a Waters Delta-Pak PrepPak C18 reverse-phase column (0.8 x 10 cm), 20 using gradient mixtures of solvent A = water/0.1% TFA and solvent B = water 10%/acetonitrile 90%, 0.09% TFA. molecular weight of the peptides was determined by electrospray mass spectrometry recorded on a triple quadrupole mass spectrometer (PE SCIEX API III), as 25 described elsewhere (Haviland et al, 1995). 1H-NMR spectra were recorded on either a Bruker ARX 500 MHz or a Varian Unity 400 spectrometer. Proton assignments were determined by 2D NMR experiments (DFCOSY, TOCSY, NOESY).

Non-peptidic compounds were synthesized using conventional organic chemical methods. Compounds were analysed by <sup>1</sup>H-NMR spectroscopy and by mass spectrometry.

#### Peptide synthesis

35 Some representative peptide syntheses are now given. Linear peptide sequences were assembled by manual step-wise solid-phase peptide synthesis with HBTU \_

activation and DIEA in situ neutralisation. Boc chemistry
was employed for temporary Nα-protection of amino acids
with two 1 min treatments with TFA for Boc group removal.
The peptides were fully deprotected and cleaved by

5 treatment with liquid HF (10 ml; p-cresol (1 ml); -5°C; 12 hr). Analytical HPLC (gradient; 0% B to 50% B over
40 min): 7, Rt = 32.0 min, [M+H]+(calc.) = 900.5,
[M+H]+(exper.) = 900.7; 8, Rt = 32.2 min,
[M+H]+(calc.) = 899.6, [M+H]+(exper.) = 899.7;

10 9, Rt = 30.0 min, [M+H]+(calc.) = 900.5, [M+H]+(exper.) =
900.7; 10, Rt = 23.8 min, [M+H]+(calc.) = 860.5,
[M+H]+(exper.) = 860.5.

Structures for the peptides are shown in Table 4 below.

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#### a) Synthesis of cycle 11

This is a general method used for the synthesis of a wide range of cyclic antagonists covered by this patent. For example, in the case of cycle 11, its linear precursor peptide was synthesised by Fmoc chemistry using HBTU/DIEA activation on an Fmoc-D-Arg(Mtr)-Wang resin. Fmoc group removal was effected using two 1 min treatments with 50% piperidine/DMF. Cleavage and deprotection using 95% TFA/2.5% TIPS/2.5% H2O gave the Mtr-protected peptide, which was purified by RP-HPLC. Cyclization of the 10 protected, purified peptide using 3eq BOP and 10eq DIEA at a 1 mM concentration in DMF stirring for 15 hr gave the cyclised product, which was fully deprotected using 1M TMSBr in TFA. A final RP-HPLC purification gave the desired peptide in yields of 50% for the cyclisation. Rt = 15 37.7 min,  $[M+H]^+$ (calc.) = 910.5,  $[M+H]^+$ (exper.) = 910.7.

#### b) Synthesis of cycle 12

Cyclization of the cleaved and fully deprotected peptide was achieved by stirring a 1mM solution in DMF with 3eq BOP and 10 eq pyridine as base for 15hr. A final RP-HPLC purification gave the desired peptide in yields of 22% for the cyclization. Rt = 37.3 min, [M+H]+(calc.) = 896.5, [M+H]+(exper.) = 896.5.

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#### NMR Structure Determination

 $1_{\rm H-NMR}$  spectra were recorded for compound 7 (3 mg in 750 µl d<sub>6</sub>-DMSO,  $\delta$  2.50) referenced to solvent on a Varian Unity 400 spectrometer at 24°C. Two dimensional  $1_{\rm H-NMR}$  NOESY (relaxation delay 2.0s, mix time 50-300 ms), DFQ- COSY and TOCSY (mixing time 75ms) experiments were acquired and recorded in phase sensitive mode. Acquisition times = 0.186 s, spectral width = 5500Hz, number of complex points (t<sub>1</sub> dimension) = 1024 for all experiments. Data was zero-filled and Fourier transformed to 1024 real points in both dimensions.

NMR data was processed using TRIAD software (Tripos Assoc.) on a Silicon Graphics Indy work station. 2D NOE cross peaks were integrated and characterised into strong (1.8-2.5Å), medium (2.3-3.5Å) and weak (3.3-5.0Å). Preliminary three-dimensional structures were calculated from upper and lower distance limit files using Diana 2.8 (69 distance constraints, including 27 for adjacent residues and 6 further away) with the redundant dihedral angle constraints (REDAC) strategy. Upper and lower distance constraints were accurately calculated using 10 MARDIGRAS. At this stage the peptide was examined for possible hydrogen bonds, and these were added as distance constraints. The 50 lowest energy Diana structures were subjected to restrained molecular dynamics (RMD) and energy minimisation (REM). Initially, REM consisted of a 50 step 15 steepest descent followed by 100 step conjugate gradient minimisation. RMD was performed by simulated heating of the structures to 300K for 1ps, followed by 500K for 1ps. The temperature was gradually lowered to 300K over 2ps and finally for 2ps at 200K. REM was performed again with a 50 20 step steepest descent, 200 step conjugate gradient followed by a 300 step Powell minimisation. The final structures were examined to obtain a mean pairwise rms difference over the backbone heavy atoms (N,  $C\alpha$  and C). Twenty of the 50 structures had a mean rmsd < 0.5 Å for all backbone atoms 25 (O, N, C).

#### Molecular Modelling

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A model of cycle 12, shown in Figure 7, was

created from the NMR structure of 7 by deleting all NMR

constraints, fusing the ornithine side chain amine to the

C-terminal carboxylate of d-Arg to form an amide, and

minimising using Powell forcefield (1000 iterations). The

modelled structure was then superimposed on the NMR

structure with an rmsd 0.224Å.

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#### Receptor-Binding Assay

Assays were performed with fresh human PMNs, isolated as previously described (Sanderson et al, 1995), using a buffer of 50 mM HEPES, 1 mM CaCl<sub>2</sub>, 5 mM MgCl<sub>2</sub>, 0.5% bovine serum albumin, 0.1% bacitracin and 100 µM phenylmethylsulfonyl fluoride (PMSF). In assays performed at 4°C, buffer, unlabelled human recombinant C5a (Sigma) or peptide, Hunter/Bolton labelled 125I-C5a (~ 20 pM) (New England Nuclear, MA) and PMNs  $(0.2 \times 10^6)$  were added sequentially to a Millipore Multiscreen assay plate (HV 10 0.45) having a final volume of 200 µL/well. After incubation for 60 min at 4°C, the samples were filtered and the plate washed once with buffer. Filters were dried, punched and counted in an LKB gamma counter. Non-specific binding was assessed by the inclusion of 1 mM peptide or 15 100 nM C5a which typically resulted in 10-15% total binding.

Data was analysed using non-linear regression and statistics with Dunnett post test.

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#### Myeloperoxidase Release

Cells were isolated as previously described (Sanderson et al, 1995) and incubated with cytochalasin B (5 $\mu$ g/mL, 15 min, 37°C). Hank's Balanced Salt solution containing 0.15% gelatin and peptide was added on to a 96 well plate (total volume 100  $\mu$ L/well), followed by 25  $\mu$ L cells (4x10<sup>6</sup>/mL). To assess the capacity of each peptide to antagonise C5a, cells were incubated for 5 min at 37°C with each peptide, followed by addition of C5a (100 nM) and further incubation for 5 min. Then 50  $\mu$ L of sodium phosphate (0.1M, pH 6.8) was added to each well, the plate was cooled to room temperature, and 25  $\mu$ L of a fresh mixture of equal volumes of dimethoxybenzidine (5.7 mg/mL) and H<sub>2</sub>O<sub>2</sub> (0.51%) was added to each well. The reaction was stopped at 10 min by addition of 2% sodium azide. Absorbances were measured at 450 nm in a Bioscan 450 plate

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reader, corrected for control values (no peptide), and analysed by non-linear regression.

## In Vivo Assays of Anti-Inflammatory Activity

The following well-known in vivo assay systems may be used to assess the anti-inflammatory activity of compounds of the invention. All assay data are analysed using non-linear regression analysis and Student's t-test, analysis of variance, with p<0.05 as the threshold level of significance.

#### (a) Carrageenan Paw Oedema

Anaesthetised (i.p. ketamine & xylazine) Wistar rats (150-200g) or mice were injected with sterilised air (20ml day 1, 10ml day 4) into the subcutaneous tissue of the back. The cavity can be used after 6 days, whereupon carrageenan (2ml, 1% w/w in 0.9% saline) was injected into the air pouch and exudate was collected after 10 hr. Test compounds are administered daily after Day 6 and their anti-inflammatory effects assayed by differential counting of cells in the air-pouch exudate. Animals were killed at appropriate times after injection and 2ml 0.9% saline was used to lavage the cavity, lavage fluids were transferred to heparinised tube and cells were counted with a haemocytometer and Diff-Quik stained cytocentrifuged preparation.

Alternatively, a routine carrageenan paw oedema was developed in Wistar rats by administering a pedal injection of carrageenan to elicit oedema which is visible in 2h and maximised in 4h. Test compounds are given 40 min before inflammagen and evaluated by microcaliper measurements of paws after 2 & 4 hr. See Fairlie, D.P. et al (1987). Also see Walker and Whitehouse (1978).

#### 35 (b) Adjuvant Arthritis.

Adjuvant arthritis was induced in rats (3 strains) either microbially (injection of heat-kilted

Mycobacterium tuberculosis) or chemically (with avridine) by inoculation with the arthritogenic adjuvant coadministered with oily vehicles (Freund's adjuvants) in the tail base. (See Whitehouse, M. W., Handbook of Animal Models for the Rheumatic Diseases, Eds. Greenwald, R. A.; Diamond, H. S.; Vol. 1, pp. 3-16, CRC Press)

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Within 13 days the adjuvant arthritis is manifested by local inflammation and ulceration in the tail, gross swelling of all four paws, inflammatory lesions in paws and ears, weight loss and fever. These symptoms, which are similar to those of inflammatory disease in humans (Winter and Nuss, 1966), can be alleviated by agents such as indomethacin or cyclosporin which also show beneficial effects in man (eg. Ward and Cloud, 1966). Without drug treatment at Day 14, arthritic rats had hypertrophy of the paws, reduced albumin but raised acute phase reaction proteins in serum, and depressed hepatic metabolism of xenobiotics as indicated by prolonged barbiturate-induced sleeping times.

To assess activity, compounds were administered for 4 days orally (\leq10mg/kg/day) or i.p. from Days 10-13 following inoculation with arthritogen (Day 0). The inflammation was either not visible or very significantly reduced in rear or front paws as assessed by microcaliper measurements of paw thickness and tail volume, as well as by gross inspection of inflammatory lesions. Animals are sacrificed by cervical dislocation on Day 18 unless arthritis signs are absent, whereupon duration of observations is continued with special permission from the Ethics committees. Experiments are staggered to maximise throughput and allow early comparisons between compounds. This routine assay is well-accepted as identifying anti-inflammatory agents for use in humans.

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# Example 1 Structure-Activity Relationship of C5a Agonists

We have focussed on the C-terminal residues of C5a, in order to explore structure-activity relationships in the search for peptide sequences with potent agonist 5 activity. Many of these peptides are full agonists relative to C5a, but have markedly lower potency (Sanderson et al, 1994, 1995; Finch et al, 1997). Our initial structure-activity investigations have been particularly informative. Mutating the decapeptide C-terminus of C5a 10 (1, C5a65-74, ISHKDMQLGR) twice with I65Y and H67F (eg. 2) led to enhancement of agonist potency by about 2 orders of These results are summarised in Table 2. Analyses of Ramachandran plots and 2D NMR spectra for compound 2 suggested that certain structural features, 15 namely a twisted "helix-like" backbone conformation for residues 65-69 and a ß-turn for residues 71-74, might be responsible for activity. These preliminary results provided some insight to structural requirements for tight 20 binding to a C5a receptor.

Table 2
Pharmacological Activity of C5a Agonist Analogues\*

| Peptide No. | Peptide                             | Fetal Artery          | PMN Enzyme                 | Binding               |
|-------------|-------------------------------------|-----------------------|----------------------------|-----------------------|
|             |                                     | EC <sub>50</sub> (µM) | Release $\mathtt{EC}_{50}$ | Affinity              |
|             |                                     |                       | (Jm)                       | IC <sub>50</sub> (µM) |
| <b>H</b>    | C5a65-74 (ISHKDMQLGR)               | >1000                 | >1000                      | >1000                 |
| 73          | YSFKDMQLGR                          | 9.6                   | 92                         | 1.3                   |
| m           | YSFKDMPLaR                          | 0.5                   | 72                         | 3.7                   |
| 7           | YSFKPMPLaR                          | 0.2                   | 4.1                        | 6.0                   |
| Ŋ           | C5a37-46-ahxYSFKPMPLaR              | 90.0                  | 5.9                        | 0.7                   |
| v           | C5a <sub>12-20</sub> -ahxYSFKPMPLaR | 0.08                  | 0.7                        | 0.07                  |
|             | C5a                                 | 0.02                  | 0.03                       | 9000.0                |

\* Finch et al, 1997

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Compounds 4, 5 and 6 in Table 2 are the highest affinity small C5a agonists so far known, with up to 25% C5a potency in human fetal artery, 5% C5a potency in human PMN enzyme release assays and 1% C5a affinity for PMN C5aR (Finch et al., 1997). For the PMN receptor, these compounds have up to 100-fold higher apparent affinity than any small molecule previously described in the literature.

The "high" affinities (70nM-6µM) of these agonist analogues for C5aR in intact PMN cells have enabled us to identify a common topographical feature in peptide agonists that correlates with expression of spasmogenic activities and enzyme-release assays in human PMNs. This preferred backbone conformation is a type II ß-turn.

The small size of these agonist peptides makes them amenable to synthetic modification to optimise their affinities, activities, and bioavailabilities, and hence useful as mechanistic probes of receptor activation.

## Example 2 NMR Structure of C5a Antagonist

We used two dimensional nuclear magnetic resonance spectroscopy to determine the three dimensional structure of 7 and found that while there is no discernible structure in water, there is evidence of a stable gammaturn structure in dimethylsulfoxide.

The 1D <sup>1</sup>H-NMR spectrum of peptide 7 in d<sub>6</sub>-DMSO at 24°C shows 4 distinct resonances for amide-NH protons, as summarized in Table 3. To establish their possible involvement in intramolecular hydrogen bonds, a deuterium exchange experiment was performed by adding a 10-fold excess of D<sub>2</sub>O to the solution. Two of the amide-NH doublets disappeared immediately, along with resonances attributable to the N-terminal methylamine protons. However, the other two amide NH resonances, as well as a broad resonance at approximately 8.05 ppm, persisted for up to 6.5 hours (Figure 2). These three slowly-exchanging protons are assigned to the amide NHs of Trp and d-Cha and the side chain amine of Lys, the slow exchange behaviour

being characteristic of hydrogen-bonding. The amine assignment was established from the TOCSY spectrumm where cross peaks were observed between the protonated amine and the  $\epsilon$ ,  $\delta$  and  $\gamma$  CH<sub>2</sub> protons. A temperature dependence study (20-60°C) of the amide-NH chemical shifts ( $\Delta\delta/T$  = 2.5 ppb/deg, dCha-NH; 6 ppb/deg, Trp-NH; 6.5 ppb, Lys-NH; 8.7ppb, Arg-NH) unambiguously confirmed the involvement of the dCha-NH only in intramolecular hydrogen bonding.

'H-NMR Assignments' for 7 in d,-DMSO Table 3

| Others  | c 7.17, 7.29; <sup>d</sup> 2.46; <sup>1</sup> 8.98 | e <sub>1.51</sub> ; t <sub>2.74</sub> , 87.76 (NH <sub>2</sub> ) | e3.61, <sup>1</sup> 3.48 | e <sub>1.43</sub> , 1.08; <sup>1</sup> 1.61, 1.58; 0.73 | c6.97,7.06,7.13,7.32,7.65; £10.80 | e3.08; £7.60 |  |
|---------|--|--|--------------------------|---|-----------------------------------|--------------|--|
| Нγ      | t  | 1.32   | 1.88, 1.78               | 0.76  | 1                                 | 1.42         |  |
| нβ      | 3.09, 3.06   | 1.74, 1.55   | 2.084, 1.74              | 1.19, 1.06  | 3.11, 2.94                        | 1.73, 1.58   |  |
| Нα      | 4.06   | 4.54   | 4.30                     | 4.35  | 4.65                              | 4.20         |  |
| NHq     | ı  | 8.83   | ı                        | 7.91  | 8.01                              | 8.44         |  |
| Residue | MePhe  | Lys  | Pro                      | d-Cha   | Trp                               | d-Arg        |  |

Referenced to residual  $d_5-DMSO$  at 2.50 ppm.

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3JNH-CaH values (Hz):7.91 (Lys), 7.77 (d-Arg), 8.34 (Trp), 8.53 (d-Cha). Amide NHs, υ

Aromatics

N-Me. Q

 $NH/NH_2$  amine.

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A series of 2D  $^{1}$ H-NMR spectra were measured for 7 at 24°C in d<sub>6</sub>-DMSO to determine the three-dimensional structure. TOCSY and DFQ-COSY experiments were used to identify residue types, while sequential assignments were made from analysis of NOESY data. From a series of 100 structures generated from NOESY data, fifty of the lowest energy structures were subjected to restrained molecular dynamics (200K-500K) and energy minimised. A set of 20 calculated structures with a root mean square deviation (rmsd) < 0.5Å (backbone atoms) are superimposed in Figure 3, and clearly depict a turn conformation.

In combination, the NMR constraint data,  $^3J_{
m NH-COH}$ values, deuterium exchange and temperature dependence data establish an unusual turn structure for hexapeptide 7 which. is constrained by up to three hydrogen bonds, as shown in Figure 4. The evidence is very strong for one intramolecular hydrogen bond from dCha-NH....OC-Lys (2.72 Å, N-H..O angle 157°, C=O...H angle 84°), forming a 7-membered ring that defines an inverse γ-turn. dChaNH-O-TrpNH angle is 56.4°. The deuterium exchange data and NMR constraint data together point to a second intramolecular hydrogen bond Trp-NH....OC-Lys (3.31 Å, N-H..O angle 159°, CO...H angle 137.3°) forming a 10-membered ring characteristic of a  $\beta$ -turn. The  $\phi$  and  $\psi$  angles  $(\phi_2 = -58.4^{\circ}, \psi_2 = 62.0^{\circ}; \phi = 96.6^{\circ}, \psi_3 = 16.6^{\circ})$  most closely match a type II β-turn (Bandekar, 1993; Hutchinson and Thornton, 1994) which is distorted by the presence of the γ-turn wholly within the  $\beta$ -turn.

To our knowledge this is the first example of an intramolecular hydrogen bond between residues within a  $\beta$ -turn, although there are many examples of hydrogen bonds between a residue within the "10 membered ring" of a  $\beta$ -turn and a residue outside of it (Bandekar, 1993). A third hydrogen bond (2.76Å, N-H...O angle 160.3°), between the side-chain amine of Lys and the C-terminal carboxylate, is suggested by the NMR constraint data, by slow NH/ND exchange and by detection of a weak NOE between Lys-

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NH...Trp- $\alpha$ CH<sub>2</sub>. This may further constrain the molecule into the observed turn conformation. Such ion-pairing is common in dipolar aprotic solvents such as dimethylsulphoxide and may also be relevant in a hydrophobic protein environment.

NMR solution structures have also been determined for several of the cyclic antagonists described in the following examples, and show that in each case the type II β-turn is preserved and stabilized by the cyclic structure.

The constraining  $\beta$  and  $\gamma$  turns proposed in the linear peptide 7 have parallels in cyclic peptides. have previously detected overlapping  $\beta$  and  $\gamma$  turns in a cyclic octapeptide from ascidiacyclamide (Abbenante et al, Combinations of a  $\beta$ - and  $\gamma$ -turn have also been found in the backbones of cyclic penta- and hexapeptides, particularly those containing alternating D- and L-amino acids (Marraud and Aubry 1996; Fairlie et al, 1995; Kessler et al, 1995; Stradley et al, 1990). For example a type II  $\beta$ -turn and an inverse  $\gamma$ -turn have been identified in cyclic 20<sup>f.</sup> antagonists c-(D- Glu-Ala-D-allo-Ile-Leu-D-Trp] (Thara et al, 1991; Coles et al, 1993; Ihara et al, 1992; Bean et al, and c-(D-Asp-Pro-D-Val-Leu-D-Trp) (Bean et al, 1994) for endothelin receptors, and in members of the rhodopsin family of G protein-coupled receptors with seven transmembrane domains (X.-M.Cheng et al, 1994). In the latter case, as in 7, an inverse  $\gamma$ -turn forms between residues (Asp-CO....Val-NH, Lys-CO....dCha-NH) that flank the proline.

Structure-Activity Relationships In Vitro 30 Example 3 We also examined the receptor-binding and antagonist activity of the hexapeptide 7 for comparison with our new compounds. The previous report by Konteatis et al (1994) concerned the ability of 7 to compete with C5a binding to receptors on isolated PMN membranes (IC50 35 70 nM), which is not necessarily physiologically relevant. We examined competition between 7 and C5a using intact PMN

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cells, and found that, under these conditions, 7 binds with much lower receptor affinity of IC<sub>50</sub> 1.8 µM. We confirmed that 7 is a full antagonist with no agonist properties. These results are summarized in Figure 5a and Table 4. The relative affinity (ratio) of 7 for the C5aR in intact PMNs in our assays was similar to that previously reported for isolated PMN membranes.

We have also found that **7** shows antagonist activity against both C5a (Figure 5b) and a C-terminal agonist decapeptide analogue **4** (YSFKPMPLaR) (Finch *et al*, 1997) of the C-terminus C5a<sub>65-74</sub>, suggesting that it acts on site 2 of the receptor. Compounds **7** and **4** have similar µM affinity for the receptor C5aR on intact polymorphonuclear leukocytes, as shown in Table 4.

A new discovery from the data in Table 4 is the linear correlation between the log of binding affinities and the log of antagonist potencies for these Site 2 antagonists (compounds 7-12, Table 4). The importance of this linear relationship is that since receptor affinity and antagonist activity are directly proportional, the experimentally simpler approach of measuring receptor binding may be used to estimate the antagonist activity for such small compounds, provided that there is no evidence of agonist activity.

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Table 4

Receptor-Binding Affinities and Antagonist Activities in Human PMNs

|              | Compound  | Receptor Affinity <sup>a</sup> | Antagonist Potency <sup>b</sup> Agonist Activity <sup>c</sup> | Agonist Activity <sup>c</sup> |
|--------------|---|--------------------------------|---|-------------------------------|
|              |   | IC <sub>50</sub> (µM)          | IC <sub>50</sub> (µM)   |                               |
| 7            | MeFKP(dCha)Wr                                     | 1.8 (15)                       | 0.085 (9)   | No                            |
| <b>&amp;</b> | $\mathtt{MeFKP}(\mathtt{dCha})\mathtt{Wr-CONH}_2$ | 14 (5)                         | 0.5 (3)   | No                            |
| o,           | MeFKP (dcha) WR                                   | 11 (5)                         | 0.7 (3)   | No                            |
| 10           | MeFKPLWR  | 144 (1)                        | >1000 (3)   | nd                            |
| 11           | Ac-F-[KP(dCha)Wr]                                 | 3.2 (40                        | 0.090 (5)   | No                            |
| 12           | Ac-F-[OP(dCha)Wr]                                 | 0.28 (6)                       | 0.012 (4)   | No                            |
| 4            | YSFKPMPLaR  | 6.0 <sup>d</sup>               | ı   | Yes                           |
| н            | C5a65-74, ISHKDMQLGR >1000                        | >1000 <sup>e</sup>             |   | 1                             |
|              | C5a   | 0.0008 (9)                     | ı   | Yes                           |

Corrected for amino acid content. Square brackets indicate cyclic portion. Number of experiments in parenthesis.

nd = not determined

 $^{\rm a}$  50% reduction in binding of  $^{125}{\rm I-C5a}$  to intact human PMNs

50% reduction in myeloperoxidase secretion from human PMNs mediated by 100 nM C5a

t Agonist activity in dose range 0.1 nM-1 mM

<sup>1</sup> Finch et al, 1997; <sup>e</sup> Kawai et al, 1991

It has previously been proposed that the Cterminus of C5a and of agonist peptides is essential for activity, due to its interaction with a positively-charged Arg206 of the receptor (DeMartino et al, 1995). We confirm here that the C-terminal carboxylate is indeed important 5 for activity (8 vs. 7), but wondered whether the origin of this effect might be due to hydrogen bonding between the carboxylate anion and the positively charged amine side chain of Lys. Conversion to the amide (8) certainly reduces both receptor-affinity and antagonist activity 10 approximately 5-fold. Changing chirality of the Arg-C $\alpha$  (9 vs. 7) causes a similar reduction in activity, and replacing dCha with the less bulky Leu residue (10) is also detrimental to receptor binding. However, potency is recovered for cyclic compounds 11 and 12, in which an amide 15 bond is tolerated at the C-terminus, consistent with the structural interpretation above that the advantage of the carboxylate in 7 may be associated with intramolecular hydrogen bonding. The replacement of this hydrogen bond in 7 with a covalent amide bond in 11 and 12 more effectively 20 stabilizes the turn conformation.

potency in vitro on human PMNs for compounds 15 and 17 with those for compound 7. Both 15 and 17 are potent inhibitors at nM concentrations of the action of C5a and the binding of <sup>125</sup>I-C5a to its receptor (e.g. 4, K<sub>b</sub> = 1.4nM). Their cyclic nature and the acetylation at the N-terminal phenylalanine both protect against the proteolytic degradation typically encountered by peptides, making such cyclic compounds more suitable than acyclic peptides as drug candidates. The results are shown in Table 5.

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( )

Table 5

Receptor Binding and Antagonist Activity

of Cyclic Molecules

Isomer Receptor Agonist Compound n  $\mathbf{R}$ Activity Affinity μм 9 No s-13 1 Н R-34 No 14 0.3 15 Н s-No 2 3.7 R-No 16 0.3 No s-17 3 Ac 38 No R-11 Аc 3.2 No 18 S-4 Ac 51 No 12 R-Ac

Refers to stereochemistry of Arg side chain.

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## Example 4 Cyclic Antagonists of C5a

Some examples of these cyclic antagonists and their apparent receptor-binding affinities and antagonist potencies are given in Tables 4, 5 and 6 as well as in Figures 5 and 6.

Effect of Cyclisation on Antagonist Binding Affinity and Antagonist Potency Table 6

|     | PEPTIDE                      | pD2 ± SE²  | IC50     | (u)     | pD2 ± SE <sup>b</sup> | IC50     | (n) |
|-----|------------------------------|--|----------|---------|-----------------------|----------|-----|
|     |                              |  | (Mt)     |         |                       | q (۱۳۲۱) |     |
| 급   | AcF-[KPdChaWr]               | 5.49 ± 0.22  | 3.2      | 4       | 7.07 ± 0.29           | 0.09     | 2   |
| 18  | AcF-[OPdChawr]               | 6.44 ± 0.14*   | 0.4      | თ       | $7.30 \pm 0.09$       | 0.05     | σ   |
| 19  | [FWPdChaWr]                  | 4.37 ± 0.36*   | 43       | е       | nd                    |          |     |
| 20  | AcF-[KMdChaWr]               | 4.81 ± 0.06  | 15       | 2       | nd                    |          |     |
| 21  | AcF-[KKdChaWr]               | 3,94 ± 0.4   | 116      | м       | 4.88                  | 13       | н   |
|     |                              | ,  |          | •       | 4                     |          | į   |
| Eff | Effect of length of li       | inker in cycle on antagonist binding affinity and antagonist | antagoni | st bind | ing attinity and      | ancagoni | នក  |
| pot | potency                      |  |          |         |                       |          |     |
| 22  | AcF-[XPdChaWr]               | $5.02 \pm 0.07$  | 9.5      | m       | $4.71 \pm 0.23$       | 20       | m   |
| 23  | AcF-[X <sup>2</sup> PdChaWr] | 4.77 ± 0.14*   | 17       | м       | 6.09 ± 0.08*          | 8.0      | 4   |
| 11  | AcF-[OPdChaWr]               | 4.60±0.06*   | 16       | 4       | $6.42 \pm 0.10$       | 0.4      | 4   |
| 24  | AcKF-[OPdChaWr]              | $4.96 \pm 0.03$  | 11       | က       | 6.73                  | 0.2      | 1   |

( )

Table 6 (cont.)

|             | GUTHOOG                | 20 + CDa         | T<br>L   | (11) | กไว + ฉูย <sup>b</sup> | TCEN            | (n |
|-------------|------------------------|------------------|----------|------|------------------------|-----------------|----|
|             | 7 EF 1 1 D E           | ) 1<br>1<br>1    | в (Мц)   | (11) |                        | وركير<br>(بلال) | () |
| 14          | F-[XPdChaWr]           | 4.39 ± 0.10*     | 41       | 3    | pu                     | !               |    |
| 16          | $F-[X^2$ PdCha $Wr]$   | $5.42 \pm 0.05$  | 3.8      | 3    | $6.70 \pm 0.04$        | 0.4             | 3  |
| 25          | F-[OPdChaWr]           | $5.51 \pm 0.07$  | 3.1      | က    | 5.79 ± 0.34*           | 1.6             | т  |
| 56          | F-[KPdChaWr]           | $5.09 \pm 0.08$  | 8.1      | ю    | 5.55 ± 0.57*           | 2.8             | Э  |
| n<br>f<br>f | Rffect of I Arg on ant | tagonist binding | affinitv |      | and antagonist         |                 |    |
| notency     |                        |                  |          |      |                        |                 |    |
| 17          | ACF-[OPdChawR]         | 6.57 ± 0.05*     | 0.3      | m    | 7.91 ± 0.17*           | 0.01            | m  |
| 13          | F-[XPdChaWR]           | +                | 10       | ю    | 5.63 ± 0.13*           | 2.4             | m  |
| 15          | $F - [X^2 PdChaWR]$    | $6.50 \pm 0.04*$ | 0.3      | Ŋ    | $7.36 \pm 0.13$        | 0.04            | т  |
| 27          | F-[OPdChaWR]           | $7.21 \pm 0.01*$ | 90.0     | ო    | $7.41 \pm 0.14$        | 0.04            | m  |
| 8           | F-[KPdChaWR]           | $6.50 \pm 0.12*$ | 0.3      | 4    | 6.69 ± 0.04            | 0.2             | 3  |

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- a  $pD_2/IC_{50}$ ; concentration of peptide resulting in 50% inhibition in the binding of [ $^{125}I$ ]C5a to intact PMNs. The IC50 is the antilog of the mean pD2 value
- b  $pD_2/IC_{50}$ ; concentration of peptide resulting in 50% inhibition in the ability of C5a (100 nM )to cause the release of MPO from PMNs

 $X = (CH_2) - NH_2$ 

 $X^2 = (CH_2)_2 - NH_2$ 

pD2 values are expressed as mean ± SE

- 10 n represents the number of experiments performed
  - \* Significant change in affinity/potency compared to NMeFKPdChaWr (p<0.05)
  - # indicates isomer number

These results demonstrate:

- (1) that the cyclic molecules have higher apparent receptoraffinity and may be more potent antagonists than acyclic (linear) peptides,
- (2) that one of the two possible cyclic

  20 diastereomers is consistently favoured for binding to the

  C5a receptor, and it is surprisingly the opposite

  stereochemistry (L-arginine) to that favoured in the linear

  compounds (D-arginine)
- (3) that the cycles have an optimum ring size 25 for receptor-binding,
  - (4) that there is a pseudo-linear relationship between log (antagonist potency) and log (receptor affinity).

of some examples of cyclic antagonists of C5a, and their ability to bind to, and inhibit, binding of C5a to human PMNs is illustrated in Figure 6. Surprisingly these data show that the L-arginine is preferred over the D-arginine, in contrast to the linear compound 7 in which the D-arginine confers higher affinity for the receptor than does L-arginine. The data also show that the size of the macrocycle is optimal when n = 2 or 3, the smaller cycle

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where n=1 and the larger cycle when n=4 being clearly less active. This requirement for a tightly constrained cycle is probably due to the need to correctly position the attached side chain residues of, for example, Trp, dCha, Arg and Phe for interaction with the receptor.

Computer Modelling of Antagonist Structures

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Example 5

Table 2).

of the cyclic antagonist 12 with the NMR solution structure for the acyclic antagonist 7. These backbone structures are strikingly similar, and strongly suggest that the receptor-binding conformations of these molecules involve the same turn structure. Compound 12, a more potent antagonist than 11, also has a shorter linker, which tightens the turn and slightly alters the conformational space accessible to the key side chains of Phe, dCha, Trp and Arg. The conformational limitations placed on the hexapeptide derivative 12 by the cycle are responsible for

a ≥104 increase in receptor-binding affinity over the

There is a correlation between binding affinities and antagonist potency for the site 2 antagonists (compounds 7-12, Table 2). It thus appears that antagonist potency is dependent upon changes that occur at site 2 alone. Without wishing to be bound by any proposed mechanism, we believe that this may be because the mechanism of antagonism is related to conformational change to a turn conformation induced by 7 at site 2 of the receptor.

conformationally flexible decapeptide C-terminus of C5a (1,

Example 6 Characterisation of C5aRs on Different Cells

Currently there is no information about different
types of C5aRs. We have previously shown marked
differences in the responsiveness of different cells
containing functional C5aRs to agonists (Sanderson et al,
1994, 1995; Finch et al, 1997) and we can now provide more

\* WO.99/00406 PCT/AU98/00490

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information by examining potency and efficacy of selective agonists and antagonists relative to human recombinant C5a. For agonists, the tissue or cell selectivity may reveal functionally different receptors. Binding assays using human PMNs, U937 cells, or circulating monocytes are used to determine affinities for C5aRs. Selectivity for different C5aRs is ascertained by differential antagonism. This combined approach allows pharmacological characterisation of new agonists or antagonists, and may lead to a potential functional classification of C5aRson different cells.

Example 7(a) Neutropenia and C5a Antagonism In Vivo

Compounds were evaluated in an acute model of

C5a-induced neutropenia. Transient neutropenia maximises 5
min after i.v. C5a and is profound, with >90% of
circulating neutrophils disappearing from circulation at
effective doses of C5a, as shown in Figure 8. The
neutropenia is due to transient adherence of circulating

neutrophils to the vascular endothelium. Preliminary
data show that neutropenia caused by i.v. C5a is blocked
by a C5a antagonist. For example, F-[OPdChaWR], (1 mg/kg),
given prior to 2µg C5a i.v., inhibits C5a-induced
neutropenia in vivo (Figure 8).

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## Example 7(b) Inhibition of Lipopolysaccharide-induced Effects by C5a Antagonists

LPS causes rapid neutropenia in rats. If this effect of LPS is blocked by C5a antagonists, then C5a may be of major importance in the acute effects of LPS, and the results shown in Figure 9 were in agreement with this hypothesis. C5a antagonists were injected (bolus i.v.) 10 min prior to challenge with LPS. Rats were anaesthetised, and blood samples (0.3ml) were taken for measurements of PMNs. PMNs are isolated and quantified. Preliminary results show that F-[OPdChaWR], (1 mg/kg), given prior to i.v.LPS, inhibits neutropenia..

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The results also indicate that the C5a antagonist inhibits the increase in haematocrit caused by LPS, showing that vascular leakage of serum caused by LPS is also inhibited.

These results demonstrate that C5a receptor antagonists, such as those described in this invention, may have therapeutic utility in septicaemic individuals. The ability to inhibit the adherence of PMNs to vascular endothelium, and to inhibit the vascular leakage to LPS as shown by the reduction of haematocrit values, indicates powerful anti-inflammatory effects of these compounds against proinflammatory stimuli activating the complement system, such as endotoxin or LPS.

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In Vivo Activity of Cyclic C5a Antagonists 15 Example 8 Preliminary experiments in rats have revealed that the cyclic antagonists summarized in Table 5 are active at less than 20 mg/kg as anti-inflammatory agents in suppressing the onset of either carrageenan-induced paw oedema or adjuvant-induced polyarthritis. The maximally 20 effective dosages for even moderately-effective antagonists are 10 mg/kg or less, given i.p. or p.o. Many antiinflammatory drugs currently used in humans were initially evaluated in such assays, and also showed activity in these rat models of inflammation. These preliminary indications 25 of efficacy in vivo indicate that C5a antagonists have therapeutic potential in human inflammatory conditions.

Using the rat carageenan paw oedema assay, we found that a compound, AcF-[O-P-dCha-W-r], which is 100 times less active than 17 in vitro as a C5a antagonist in PMNs, has some in vivo activity in rats given 1 mg/kg of the compound I.P, 30 min prior to the carageenan injection. Paw swelling was measured for up to 4.5 hr. The results, shown in Figure 10, suggest that even this weak C5a antagonist significantly inhibits development of the oedema after 180 and 270 min. This anti-inflammatory activity suggests that C5a receptor antagonists, such as those

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described in this invention, may have therapeutic activity in diseases involving vascular leakage following inflammatory stimuli.

In recent years there have been many attempts to mimic  $\beta$ - and  $\gamma$ - turn peptides that represent bioactive 5 protein surfaces, resulting in notable mimetics for RGD (arginine-glycine-aspartate) peptides, somatostatin and opioid peptides, to name a few derived through structureactivity relationships (see for example Marraud and Aubry, 1996; Fairlie et al, 1995). Most of these examples 10 preserve a turn structure through cyclisation of the peptide. On the other hand, there are comparatively few short acyclic peptides that have been found to have substantial turn structure in solution (Dyson et al, 1988; Rizo and Gierasch, 1992; Pràcheur et al, 1994). 15 usually argued that short acyclic peptides adopt a myriad of solution structures that may include small populations of turn structures that are responsible for bioactivity.

This invention describes a series of conformationally-constrained turn-containing molecules that are preorganized for binding to the same G protein-coupled receptor(s) of human cells that are targeted by human C5a. The invention is applicable to other G protein-coupled receptors.

The principal feature of the compounds of the invention is the preorganized arrangement, which brings at least three hydrophobic groups and a charged group into neighbouring space, creating a hydrophobic surface 'patch'. These results enable the design and development of even more potent conformationally-constrained, small molecule antagonists of C5a.

In the light of the aforementioned prior art, it was surprising to find that a C-terminal carboxylate was not necessary in our compounds in order to obtain good receptor-binding or antagonist activity. The cyclic antagonists have an amide bond at the 'C-terminal' arginine position. The replacement of the carboxylate in 7 with a

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covalent amide bond effectively stabilises the required turn conformation.

Cyclic and non-peptidic antagonists have several important advantages over peptides as drugs. The cycles 5 described in this invention are stable to proteolytic degradation for at least several hours at 37°C in human blood or plasma, or in human or rat gastric juices or in the presence of digestive enzymes such as pepsin, trypsin and chymotrypsin. In contrast, short peptides composed of 10 L-amino acids are rapidly degraded to their component amino acids within a few minutes under these conditions. A second advantage lies in the constrained single conformations adopted by the cyclic and non-peptidic molecules, whereas acyclic or linear peptides are flexible 15 enough to adopt several structures in solution other than the required receptor-binding structure. Thirdly, cyclic and non-peptidic compounds such as those described in this invention are usually more lipid-soluble and more pharmacologically bioavailable as drugs than peptides, 20 which can rarely be administered orally. Fourthly, the plasma half-lives of cyclic and non-peptidic molecules are usually longer than those of peptides.

It will be apparent to the person skilled in the art that while the invention has been described in some detail for the purposes of clarity and understanding, various modifications and alterations to the embodiments and methods described herein may be made without departing from the scope of the inventive concept disclosed in this specification.

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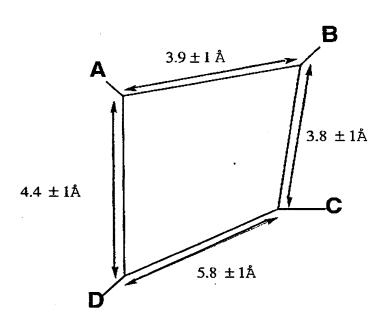
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#### CLAIMS:

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1. A compound which is an antagonist of a G proteincoupled receptor, which has no agonist activity, and which has a structure adapted to provide a framework of approximate dimensions as set out in Structure I:

### Structure I



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where the numerals refer to distances between  $C_{\alpha}$  carbons of amino acids or their analogues or derivatives, and A, B, C and D are not necessarily on adjacent amino acids, or analogues or derivatives thereof; and

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where the critical amino acid side chains are designated by A, B, C and D, or are as defined below;

A is any common or uncommon, basic, charged amino acid side chain which serves to position a positively charged group in this position;

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B is any common or uncommon, aromatic amino acid side chain which serves to position an aromatic side-chain in this position;

C is any common or uncommon, hydrophobic amino acid side chain which serves to position any alkyl, aromatic or other group in this position;

D is any common or uncommon, aromatic amino acid which serve to position an aromatic side-chain in this position, and has the structure:

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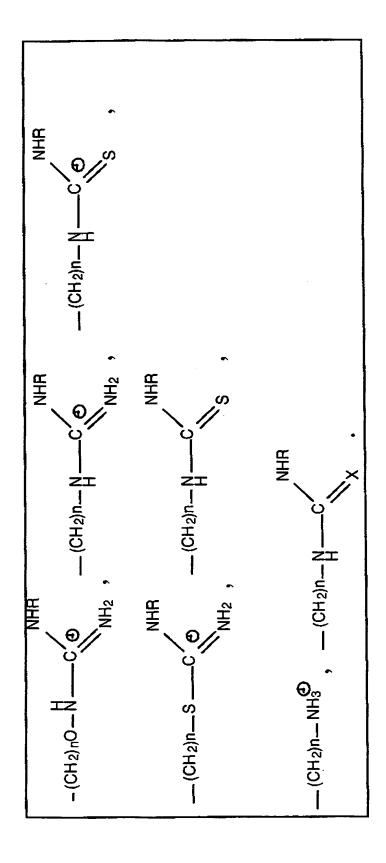
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where Z is indole, indole methyl, benzyl, benzene, naphthyl, naphthyl methyl, or a derivative thereof; and R is H or an alkyl, aromatic, acyl or aromaticacyl group.

- 2. An antagonist according to Claim 1, in which the G protein-coupled receptor is the C5a receptor.
- 3. An antagonist according to Claim 1 or Claim 2, in 20 which

A is one of the following side-chains



where

or another mimetic of an arginine side chain;

X is H, NCN, NNO2, CHNO2 or NSO2NH2;

n is an integer from 1 to 4, and

5 R is H or an alkyl, aryl, CN, NH2 or OH;

B is an indole, indole methyl, benzyl, phenyl, naphthyl, naphthyl methyl, cinnamyl group, or any other derivative of the aromatic group;

C is D- or L-cyclohexyl alanine (Cha), leucine, valine, isoleucine, phenylalanine, tryptophan, methionine; and

R is methyl, ethyl, propyl, butyl,  $-CO-CH_2CH_3$ ,  $-CO-CH_3$ ,  $-CO-CH_2CH_3$ ,  $-CO-CH_2CH_3$ ,  $-CO-CH_2Ph$ , or -CO-Ph.

- 4. An antagonist according to any one of Claims 1 to
- 15 3, which is linear, and comprises a type II  $\beta$ -turn.
  - 5. An antagonist according to Claim 4, in which the type II  $\beta$ -turn comprises a  $\gamma$ -turn within the type II  $\beta$ -turn.
- 6. An antagonist according to any one of Claims 1 to 20 5, of formula

Ac-F-[KP(dCha)Wr], or Ac-F-[OP(dCha)Wr]

- 7. An antagonist according to any one of Claims 1 to 6 in which A is L-arginine.
- 25 8. An antagonist according to Claim 1, which has antagonist activity against C5aR, has no agonist activity against C5a, and has the general formula:

Structure II

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where A is H, alkyl, aryl,  $NH_2$ , NHalkyl,  $N(alkyl)_2$ , NHaryl or NHacyl;

B is an alkyl, aryl, phenyl, benzyl, naphthyl or indole group, or the side chain of a D- or L-amino acid selected from the group consisting of phenylalanine, homophenylalanine, tryptophan, homotryptophan, tyrosine, and homotyrosine;

C is the side chain of a D-, L- or homo-amino acid selected from the group consisting of proline, alanine, leucine, valine, isoleucine, arginine, histidine, aspartate, glutamate, glutamine, asparagine, lysine, tyrosine, phenylalanine, cyclohexylalanine, norleucine, tryptophan, cysteine and methionine;

D is the side chain of a D- or L-amino acid selected from the group consisting of cyclohexylalanine, homocyclohexylalanine, leucine, norleucine, homonorleucine and tryptophan;

E is the side chain of a D- or L-amino acid selected from the group consisting of tryptophan and homotryptophan;

F is the side chain of a D- or L-amino acid selected from the group consisting of arginine,

25 homoarginine, lysine and homolysine; and

X is  $-(CH_2)_nNH-$  or  $(CH_2)_n-S-$ , where n is an integer of from 1 to 4,  $-(CH_2)_2O-$ ,  $-(CH_2)_3O-$ ,  $-(CH_2)_3-$ , -

 $(CH_2)_4$ -, or  $-CH_2COCHRNH$ -, where R is the side chain of any common or uncommon amino acid.

- 9. An antagonist according to Claim 8, in which F is a L-amino acid.
- 5 10. An antagonist according to Claim 9, in which F is L-arginine.
  - 11. An antagonist according to any one of Claims 8 to 10, selected from the group consisting of compounds 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27 and 28.
  - 12. An antagonist according to any one of Claims 8 to 11, in which n is 2 or 3.
  - 13. A compound which is an agonist of a G protein-coupled receptor, and which has structure III

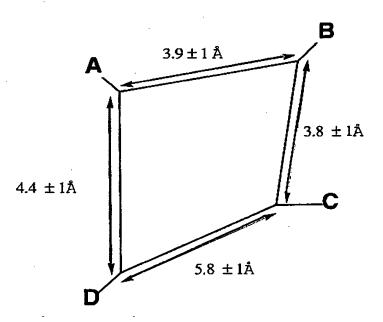
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#### Structure III



where the numerals refer to distances between  $C_{\alpha}$  carbons of amino acids or their analogues or derivatives, and A, B, C and D are not necessarily on adjacent amino acids, or analogues or derivatives thereof; and

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where B is a non-aromatic amino acid, and A, C and D are as defined in Claim 1.

- 14. A compound according to Claim 13, where B is the D- or L-form of alanine, leucine, valine, norleucine,
- 5 glutamic acid, aspartic acid, methionine, cysteine, isoleucine, serine or threonine.
  - 15. A compound according to Claim 13 or Claim 14, in which the compound is of structure IV,

10 Structure IV

where E is any amino acid other than tryptophan and homotryptophan.

16. A compound according to any one of Claims 13 to

15, wherein the compound is an agonist of C5a.

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17. A compound according to Claim 8, of structure

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- 18. A composition comprising a compound according to any one of Claims 1 to 17, together with a pharmaceutically-acceptable carrier or excipient.
- 19. A method of treatment of a pathological condition
  5 mediated by a G protein-coupled receptor, comprising the
  step of administering an effective amount of a compound
  according to any one of Claims 1 to 17, to a mammal in need
  of such treatment.
- 20. A method according to Claim 19, wherein the
  10 condition mediated by a G protein-coupled receptor involves
  overexpression or underregulation of C5a.
  - 21. A method according to Claim 19, wherein the condition is selected from the group consisting of rheumatoid arthritis, adult respiratory distress syndrome (ARDS), systemic lupus erythematosus, tissue graft
- 15 (ARDS), systemic lupus erythematosus, tissue graft rejection, ischaemic heart disease, reperfusion injury, septic shock, psoriasis, gingivitis, atherosclerosis, Alzheimer's disease, multiple sclerosis, lung injury and extracorporeal post-dialysis syndrome.
- 20 22. Use of a compound according to any one of Claims 1 to 17 in treatment of a pathological condition mediated by G protein-coupled receptors.
  - 23. Use according to Claim 22, in which the condition is mediated by C5a.

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- 24. Use according to Claim 23, in which the condition mediated by G protein-coupled receptors involves overexpression or underregulation of C5a.
- 25. Use according to any one of Claims 22 to 24, in which the condition is selected from the group consisting of rheumatoid arthritis, adult respiratory distress syndrome (ARDS), systemic lupus erythematosus, tissue graft rejection, ischaemic heart disease, reperfusion injury, septic shock, psoriasis, gingivitis, atherosclerosis,
- 10 Alzheimer's disease, multiple sclerosis, lung injury and extracorporeal post-dialysis syndrome
  - 26. Use of a compound according to any one of Claims 1 to 17 in the manufacture of a medicament for the treatment of a condition mediated by a G protein-coupled receptor.
  - 27. Use according to Claim 26, in which the condition is mediated by C5a.
  - 28. Use according to Claim 27, in which the condition mediated by G protein-coupled receptors involves
- 20 overexpression or underregulation of C5a.

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- 29. A compound according to Claim 1, substantially as hereinbefore described with reference to the examples and drawings.
- 30. A method according to Claim 19, substantially as25 hereinbefore defined with reference to the examples and drawings.

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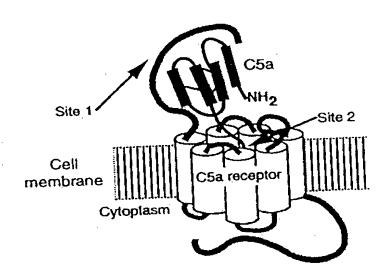


Fig. 1

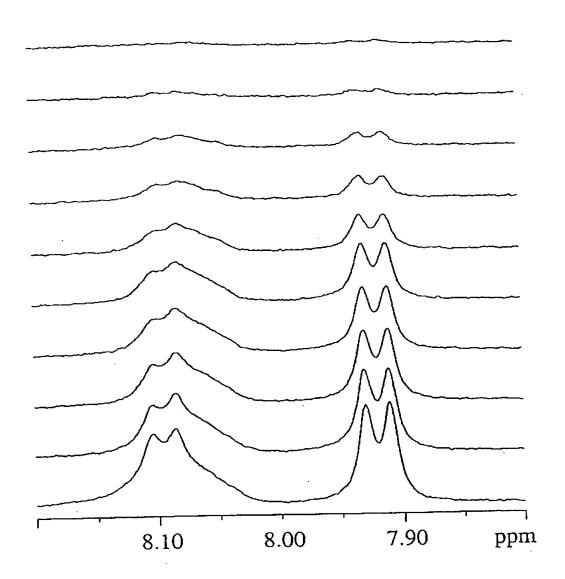


Fig. 2

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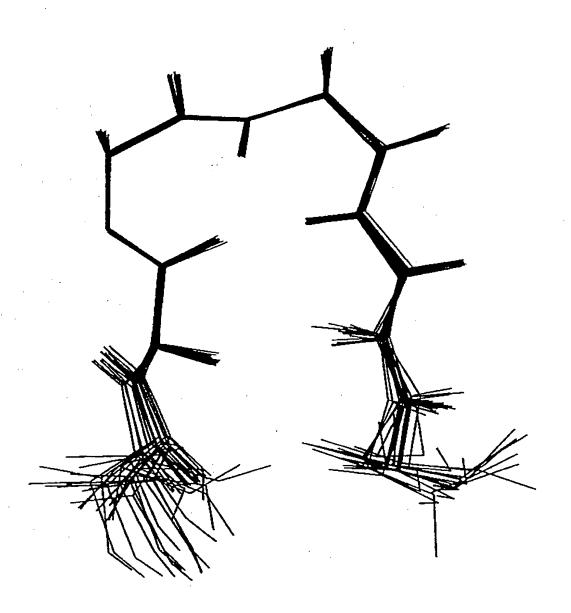
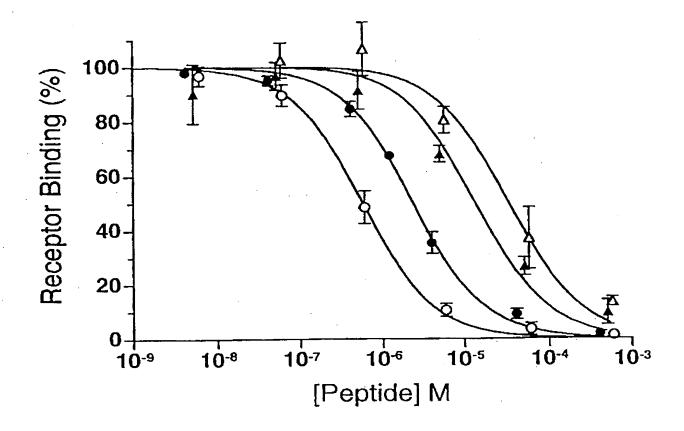


Fig. 3



Jig. 5a

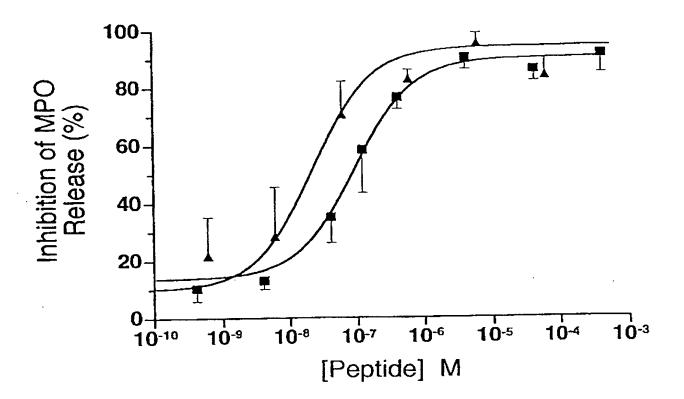
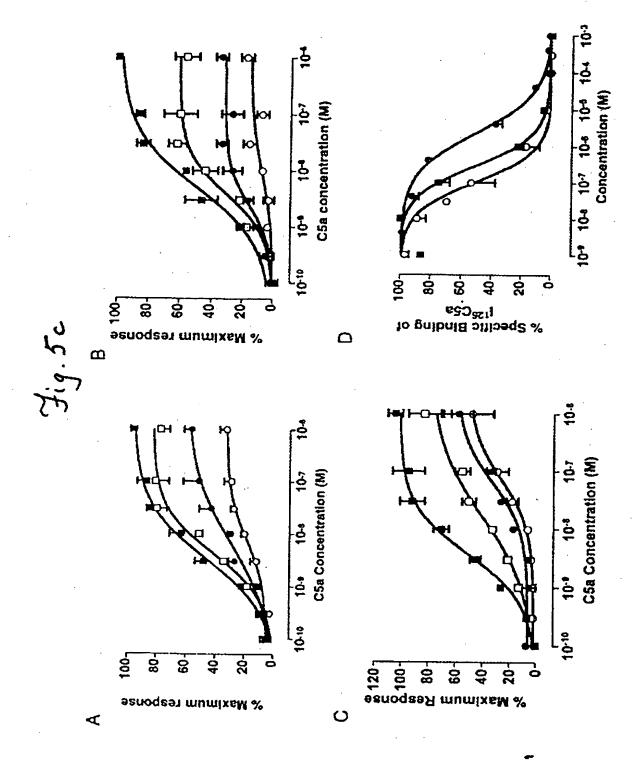


Fig. 5 b



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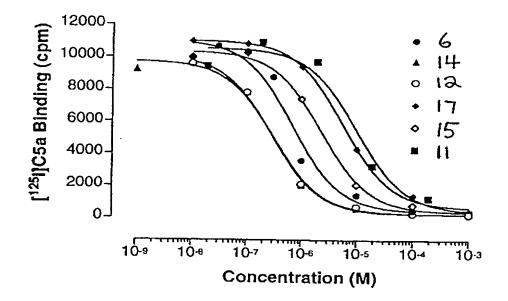


Fig. 6

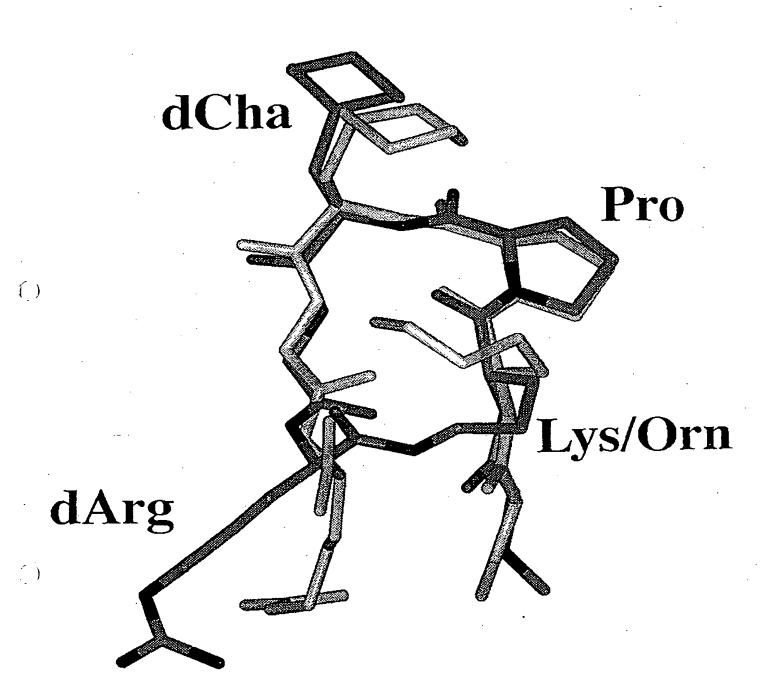


Fig. 7

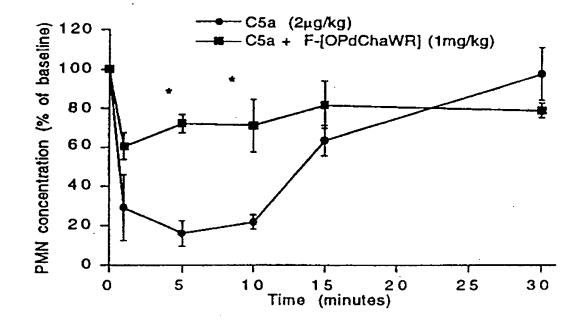
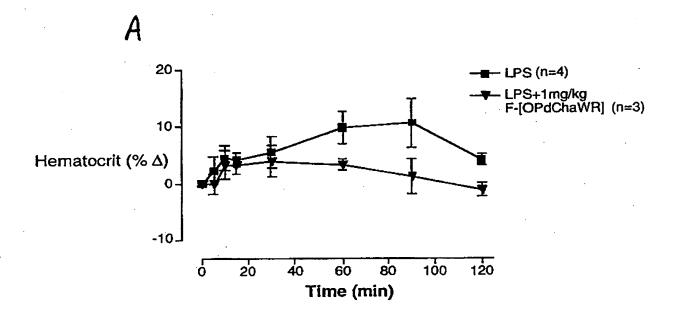


Fig. 8

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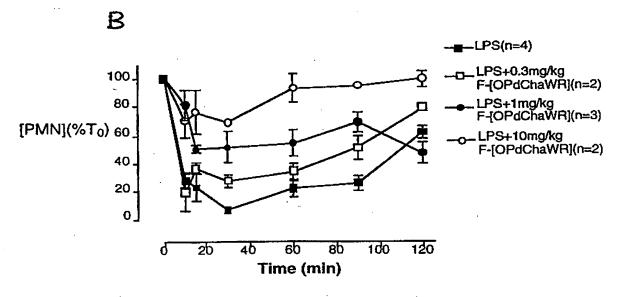
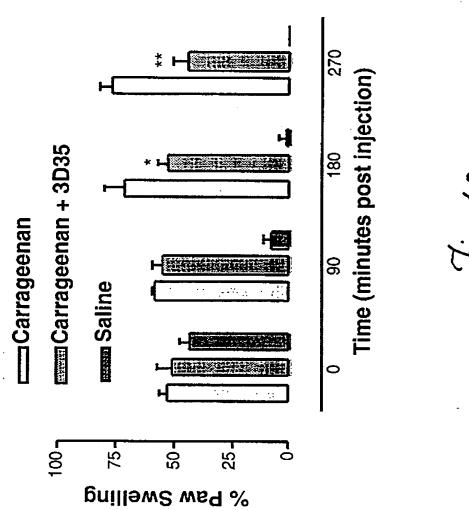


Fig. 9



Jig. 10

International Application No. PCT/AU 98/00490

| A.   | CLASSIFICATION OF SUBJECT MATTER  |   |                            |
|--|---|---|----------------------------|
| Int Cl <sup>6</sup> :  | C07K 7/06, 7/64; A61K 38/08.  |   |                            |
| According to   | International Patent Classification (IPC) or to both  | national classification and IPC                       |                            |
| В.   | FIELDS SEARCHED   |   |                            |
| Minimum docu   | umentation searched (classification system followed by ch   | assification symbols)                                 |                            |
| Documentation  | n searched other than minimum documentation to the exte   | ent that such documents are included in t             | he fields searched         |
| Electronic data<br>Medline (C5   | a base consulted during the international search (name of a and antagonist or agonist) **see continuation   | data base and, where practicable, search on last page | terms used)                |
| C.   | DOCUMENTS CONSIDERED TO BE RELEVANT   |   |                            |
| Category*  | Citation of document, with indication, where app  | ropriate, of the relevant passages                    | Relevant to claim No.      |
| P. X   | JOURNAL OF PEPTIDE RESEARCH, volume 5<br>'NMR Analysis of a Potent Decapeptide Agonist pages 226-234<br>(see the whole document, in particular page 226, Conclusions) | of Human C5a Anaphylatoxin'.                          | 1-5, 7-10,<br>12-16, 18-30 |
| JOURNAL OF IMMUNOLOGY, volume 158, February 1997, R.M. Tempero et al., 'Molecular Adjuvant Effects of a Conformationally Biased Agonist of Human C5a Anaphylatoxin', pages 1377-1382 (see in particular page 1379, Table I and page 1381, Table II)  |   |   |                            |
| X Further documents are listed in the continuation of Box C See patent family annex  |   |   |                            |
| "T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone document or other special reason (as specified)  "O" document referring to an oral disclosure, use, exhibition or other means "P" document published prior to the international filing "T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combined with one or more other such documents, such combination being obvious to a person skilled in the art document member of the same patent family |   |   |                            |
| Date of the ac   | but later than the priority date claimed  | Date of mailing of the international sea              | rch report                 |
| 22 July 1998  Name and mailing address of the ISA/AU AUSTRALIAN PATENT OFFICE PO BOX 200 WODEN ACT 2606 AUSTRALIA Facsimile No.: (02) 6285 3929  Authorized officer  GILLIAN JENKINS  Telephone No.: (02) 6283 2252  |   |   |                            |

International Application No.

PCT/AU 98/00490

| C (Continua | tion) DOCUMENTS CONSIDERED TO BE RELEVANT  |                              |
|-------------|--|------------------------------|
| Category*   | Citation of document, with indication, where appropriate, of the relevant passages   | Relevant to claim No.        |
| Х           | JOURNAL OF IMMUNOLOGY, volume 157, November 1996, L. Baranyi et al., 'Antisense Homology Boxes in C5a Receptor and C5a Anaphylatoxin', pages 4591-4601 (see in particular page 4592, column 1, Peptide Synthesis)  | 1-3, 7, 13, 14,<br>16, 18-30 |
| х           | JOURNAL OF PHARMACOLOGY AND EXPERIMENTAL THERAPEUTICS, volume 278, July 1996, R. Kawatsu et al., 'Conformationally Biased Analogs of Human C5a Mediate Changes in Vascular Permeability', pages 432-440 (see in particular page 434, Table 1)  | 1-3, 7, 13, 14,<br>16, 18-30 |
| X           | IMMUNOLOGY, volume 86, September 1995, Y. Kaneko et al., 'Antagonistic Peptides Against Human Anaphylatoxin C5a', pages 149-154 (see in particular page 150, Figure 1 and page 153, Table 1)   | 1-3, 7, 13, 14,<br>16, 18-30 |
| х           | JOURNAL OF MEDICINAL CHEMISTRY, volume 34, July 1991, M. Kawai et al., 'Identification and Synthesis of a Receptor Binding Site of Human Anaphylatoxin C5a', pages 2068-2071 (see in particular page 2070, Table II, compounds 14 and 15)  | 1-3, 7, 13, 14,<br>16, 18-30 |
| X           | JOURNAL OF IMMUNOLOGY, volume 153, November 1994, Z. D. Konteatis et al., 'Development of C5a Receptor Antagonists. Differential Loss of Functional Responses', pages 4200-4205 (see in particular page 4201, Table 1)   | 1-3, 7 13, 14,<br>16, 18-30  |
| х           | JOURNAL OF MEDICINAL CHEMISTRY, volume 38, September 1995, S. D. Sanderson et al., 'Decapeptide Agonists of Human C5a: the Relationship Between Conformation and Neutrophil Response', pages 3669-3675 (see in particular page 3670, Table I)  | 1-3, 7, 13, 14,<br>16, 18-30 |
| х           | JOURNAL OF MEDICINAL CHEMISTRY, volume 40, March 1997, A. M. Finch et al., 'Biologically Active Conformer of the Effector Region of Human C5a and Modulatory Effects of N-Terminal Receptor Binding Determinants on Activity', pages 877-884 (see in particular page 878, Tables I and II)                             | 1-4, 7, 13, 14,<br>16, 18-30 |
| <b>X</b>    | JOURNAL OF MEDICINAL CHEMISTRY, volume 35, January 1992, M. Kawai et al., 'Structure-Function Studies in a Series of Carboxyl-Terminal Octapeptide Analogues of Anaphylatoxin C5a', pages 220-223 (see in particular page 221, Table II, compounds 5 and 6)  | 1-3, 7, 13, 14,<br>16, 18-30 |
| х           | BIOCHEMICAL PHARMACOLOGY, volume 45, March 1993, G. Drapeau et al., 'Synthetic C5a Receptor Agonists. Pharmacology, Metabolism and in vivo Cardiovascular and Hematologic Effects', pages 1289-1299 (see in particular page 1291, Table I)   | 1-3, 7, 13, 14,<br>16, 18-30 |
| х           | JOURNAL OF BIOLOGICAL CHEMISTRY, volume 270, July 1995, J. A. De Martino et al., 'Arginine 206 of the C5a Receptor is Critical for Ligand Recognition and Receptor Activation by C-Terminal Hexapeptide Analogs', pages 15966-15969 (see in particular page 15967, Table I, peptides 1 and 2 and page 15968, Table II) | 13, 14, 16,<br>18-30         |
| х           | PROTEIN SCIENCE, volume 6, January 1997, X. Zhang et al., 'Solution Stucture of a Unique C5a Semi-Synthetiic Antagonist: Implications in Receptor Binding', pages 65-72 (see the whole document, in particular page 66, Figure 1)  | 1-3, 7, 13, 14,<br>16, 18-30 |
| P, X        | JOURNAL OF IMMUNOLOGY, volume 160, June 1998, T. C. Pellas et al., 'Novel C5a Receptor Antagonists Regulate Neutrophil Functions in vitro and in vivo', pages 5616-5621 (see in particular the Abstract and page 5618, Tables I and II)  | 1-3, 7, 13, 14,<br>16, 18-30 |

international Application No. PCT/AU 98/00490

| Box 1     | Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)   |
|-----------|---|
| This Inte | ernational Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following   |
| ī.        | Claims Nos.:  because they relate to subject matter not required to be searched by this Authority, namely:  |
|           |   |
| 2.        | Claims Nos.: 1-7 (in part), 13-14 (in part), 16 (in part) and 18-30 (in part).  because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically: |
|           | The claims are exceedingly broad in scope and thus a complete search was not feasible for economic reasons.  Consequently the search has been restricted to those compounds described in the examples and defined by claims 8 and 15.   |
| 3.        | Claims Nos.:  because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a)  |
| Box II    | Observations where unity of invention is lacking (Continuation of item 2 of first sheet)  |
| This Inte | ernational Searching Authority found multiple inventions in this international application, as follows:   |
| 1.        | As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims   |
| 2.        | As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.  |
| 3.        | As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:  |
| 4.        | No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:  |
| Remark    | The additional search fees were accompanied by the applicant's protest.  No protest accompanied the payment of additional search fees.  |

International Application No.

|  | PCT/AU 98/00490                   |
|--|-----------------------------------|
| Box B continued  |                                   |
| FIELDS SEARCHED  |                                   |
| CA Peptide Subsequence Search  |                                   |
| 'Phe-Und-[Lys Met Pro]-Und-Trp-Arg' / '[Pro Val Ala Leu Ile Arg His Asp Asn Glu [Leu Nie Trp]-Trp-[Arg Har Lys]' / '[Pro Ala Leu Val Ile Arg His Asp Asn Glu Gln Nie Trp]-[-Trp]-[Arg Har Lys]' /sqsp and cyclic/nte and keywords (antagonist or ago | Lys Tyr Phe Nle Trp Cys Met]-(Leu |
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